1X Crimson LongAmp Tag Reaction Buffer: 60 mM Tris-So4 (pH 9.0 @ 25°C), 20 mM (NH4)2SO4, 2 mM MgSO4, 3% glycerol, 0.06% IGEPA-CAL-A-360, 0.05% Tween 20, Acid Red. Unit Definition: One unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid insoluble material in 30 minutes at 75°C. Unit Assay Conditions: 1X ThermoPol® Reaction Buffer, 200 µM dNTPs including [3H]-dTTP and 200 µg/ml activated Calf Thymus DNA. Heat Inactivation: No. Quality Control Assays: Long Amplicon PCR: Crimson LongAmp Tag DNA Polymerase is tested for the ability to amplify a 30 kb amplicon from lambda DNA and a 30 kb amplicon from human genomic DNA. PCR: The Polymerase Chain Reaction (PCR) is a powerful and sensitive technique for DNA amplification. Tag DNA Polymerase is an enzyme widely used in PCR (3). The following guidelines are provided to ensure successful PCR using New England Biolabs’ Crimson LongAmp Tag DNA Polymerase. These guidelines cover routine PCR reactions. Amplification of templates with high GC content, high secondary structure or low template concentrations may require further optimization. Reaction setup: We recommend assembling all reaction components on ice and quickly transferring the reactions to a thermocycler preheated to the denaturation temperature (94°C). Transfer PCR tubes from ice to a PCR machine with the block preheated to 94°C and begin thermocycling. General Guidelines: 1. Template: The quality of the DNA template is essential for long-range PCR amplification. Recommended amounts of DNA template for a 50 µl reaction are as follows: a) PCR reaction may be 0.05–1 µM, typically 0.1–0.5 µM. 3. Mg++ and additives: Mg++ concentration of 1.5–2.0 mM is optimal for most PCR products generated with Crimson LongAmp Tag DNA Polymerase. The final Mg++ concentration in 1X Crimson LongAmp Tag Reaction Buffer is 2 mM. This supports satisfactory amplification of most amplicons. However, Mg++ can be further optimized in 0.5 or 1.0 mM increments using MgSO4. Amplification of some difficult targets, like GC-rich sequences, may be improved with additives, such as DMSO (4) or formamide (5). 4. Deoxynucleotides: The recommended final concentration of dNTPs for long-range PCR is 300 µM of each deoxynucleotide. 5. Crimson LongAmp Tag DNA Polymerase concentration: We generally recommend using Crimson LongAmp Tag DNA Polymerase at a concentration of 100 units/ml (5 units/50 µl reaction). However, the optimal concentration of Crimson LongAmp Tag DNA Polymerase may vary in specialized applications. 6. Denaturation: An initial denaturation of 30 seconds at 94°C is sufficient for most amplicons from pure DNA templates. For difficult templates such as GC-rich sequences, a longer denaturation of 2–4 minutes at 94°C is recommended prior to PCR cycling to fully denature the template. With colony PCR, an initial 5 minute denaturation at 94°C is recommended. During thermocycling a 10–30 second denaturation at 94°C is recommended. 7. Annealing: The annealing step is typically 15–60 seconds. Annealing temperature is based on the Tm of the primer pair and is typically 45–65°C. Annealing temperatures can be optimized by doing a temperature gradient PCR starting 5°C below the calculated Tm. We recommend using NEB’s Tm Calculator, available at www.neb.com/TmCalculator to determine appropriate annealing temperatures for PCR. When primers with annealing temperatures above 60°C are used, a 2-step PCR protocol is possible (see #10).
8. Extension:
The recommended extension temperature is 65°C. Extension times are generally 50 seconds per kb. A final extension of 10 minutes at 65°C is recommended.

9. Cycle number:
Generally, 25–35 cycles yields sufficient product. Up to 45 cycles may be required to detect low-copy-number targets.

10. 2-step PCR:
When primers with annealing temperatures above 60°C are used, a 2-step thermocycling protocol is possible.

**Thermocycling Conditions for a Routine 2-Step PCR:**

<table>
<thead>
<tr>
<th>STEP</th>
<th>TEMP</th>
<th>TIME</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial Denaturation</td>
<td>94°C</td>
<td>30 seconds</td>
</tr>
<tr>
<td>30 Cycles</td>
<td>94°C</td>
<td>10–30 seconds</td>
</tr>
<tr>
<td>60–65°C</td>
<td>50 seconds/kb</td>
<td></td>
</tr>
<tr>
<td>Final Extension</td>
<td>60–65°C</td>
<td>10 minutes</td>
</tr>
<tr>
<td>Hold</td>
<td>4–10°C</td>
<td></td>
</tr>
</tbody>
</table>

11. PCR product:
The majority of the PCR products generated using Crimson LongAmp Taq DNA Polymerase contain dA overhangs at the 3′-end; therefore the PCR products can be ligated to dT/dU-overhang vectors.

**FAQs:**

1. What are the advantages or disadvantages of Crimson LongAmp Taq DNA Polymerase?
The Crimson LongAmp Taq Reaction Buffer formulation offers three convenient features. First, the 5X Crimson LongAmp Taq Reaction Buffer contains a red dye, which serves as a visual indicator of homogeneous reaction setup. Second, the 5X Crimson LongAmp Taq Reaction buffer facilitates direct loading of PCR products on a gel. Third, the trace amount of red dye in Crimson LongAmp Taq Reaction Buffer works as a tracking dye during electrophoresis.

If the PCR products will be analyzed by absorbance or fluorescence excitation, acid red, (λ_max = 510 nm) may interfere with the assays; Therefore LongAmp Taq Reaction Buffer is recommended.

2. How do I remove the dye from my PCR reactions using Crimson LongAmp Taq DNA Polymerase?
Spin Columns for PCR clean-up can be used to remove the dye.

3. What is the recommended enzyme amount when using Crimson LongAmp?
In general, we recommend 5 units of Crimson LongAmp Taq DNA Polymerase in a 50 µl PCR reaction. For amplicons < 8 kb, we recommend 1–2.5 units per 50 µl PCR reaction for higher fidelity.

4. Can the extension step be carried out at 72°C when using Crimson LongAmp?
Yes, Crimson LongAmp Taq DNA Polymerase can be used at 72°C. However, extension at 65–68°C is a better choice for most amplicons.

5. What is the extension rate when using Crimson LongAmp?
We recommend 50 seconds per kb for maximum yields. Extension rate such as 30 seconds per kb can be used for targets up to 4 kb using a 3-step PCR protocol. Shorter extension rates such as 15 seconds per kb can be used for targets up to 2 kb using a 3-step PCR protocol on a fast PCR machine.

6. What type of DNA ends result from a primer extension reaction or a PCR reaction using Crimson LongAmp Taq DNA Polymerase?
The majority of the PCR products generated using Crimson LongAmp Taq DNA Polymerase contain dA overhangs at the 3′-end; therefore the PCR products can be ligated to dT/dU-overhang vectors.

7. Why is the product a smear when visualized on an agarose gel?
When PCR conditions are not optimal, a smear or high level of background is often observed. Try one or more of the following suggestions:
- use lower amount of enzymes
- use 65°C for extension
- raise annealing temperature
- try 2-step cycling protocols

8. Can Crimson LongAmp Taq DNA Polymerase be used to amplify GC-rich amplicons?
Yes. The addition of DMSO up to 10% helps amplify GC-rich amplicons.

**References:**


**Companion Products Sold Separately:**

LongAmp Taq (Mg-free) Reaction Buffer Pack #B0322S 6.0 ml
LongAmp Taq Reaction Buffer Pack #B0323S 6.0 ml
Crimson LongAmp Taq Reaction Buffer Pack #B0326S 6.0 ml
Magnesium Sulfate (MgSO₄) Solution #B1003S 6.0 ml
Diluent F #B8006S 4.0 ml
LongAmp Taq PCR Kit #E5200S 100 Reactions
LongAmp Taq 2X Master Mix #M0287S 100 Reactions #M0287L 500 Reactions
LongAmp Taq DNA Polymerase #M0323S 500 units #M0323L 2,500 units
Deoxynucleotide Solution Set #N0446S 25 µmol of each
Deoxynucleotide Solution Mix #N0447S 8 µmol of each #N0447L 40 µmol of each

**Limited Label License:** This product is covered by certain patents outside the United States (the ‘Non-US Patents’) corresponding to the expired U.S. Patent No. 5,436,149 (claims 6–16). The purchase of this product conveys to the purchaser only the limited, non-transferable right under the Non-US Patents to use the purchased quantity of the product for the purchaser’s own research by the purchaser only. No rights are granted to the purchaser with respect to the Non-US Patents to sell, modify for resale or otherwise transfer this product, either alone or as a component of another product, to any third party. Takara Bio reserves all other rights to the Non-US Patents, and this product may not be used in any manner other than as provided herein. For information on obtaining a license with respect to the Non-US Patents to use this product for purposes other than research, please contact Takara Bio Inc., Seta 3-4-1, Otsu, Shiga 520-2193, Japan (Fax +81-77-543-9254).

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