

# Antarctic Phosphatase



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M0289S 022140616061

## M0289S



**1,000 units**    **5,000 U/ml**    **Lot: 0221406**  
**RECOMBINANT**    **Store at -20°C**    **Exp: 6/16**

**Description:** Antarctic Phosphatase catalyzes the removal of 5' phosphate groups from DNA and RNA. Since phosphatase-treated fragments lack the 5' phosphoryl termini required by ligases, they cannot self-ligate (1). This property can be used to decrease the vector background in cloning strategies.

**Heat Inactivated In 5 Minutes at 70°C**

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**Source:** An *E. coli* strain that carries the TAB5 AP gene, originally cloned in plasmid pNI (2), recloned in plasmid pEGTAB7-4.1(3).

### Applications:

- Removing 5' phosphates from DNA, RNA, rNTPs and dNTPs
- Preparation of templates for 5' end labeling
- Prevention of recircularization of cloning vectors
- Dephosphorylation of proteins
- Removal of dNTPs and pyrophosphate from PCR reactions

Supplied in: 10 mM Tris-HCl (pH 7.4), 1 mM MgCl<sub>2</sub>, 0.01 mM ZnCl<sub>2</sub>, 1 mM DTT and 50% glycerol.

### Reagents Supplied with Enzyme:

10X Antarctic Phosphatase Reaction Buffer.

**Reaction Conditions:** 1X Antarctic Phosphatase Reaction Buffer. Incubate at 37°C.

### 1X Antarctic Phosphatase Reaction Buffer:

50 mM Bis-Tris-Propane HCl

1 mM MgCl<sub>2</sub>

0.1 mM ZnCl<sub>2</sub>

pH 6.0 @ 25°C

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**Unit Definition:** One unit is defined as the amount of enzyme that will dephosphorylate 1 µg of pUC19 vector DNA cut with HindIII (5' protruding ends), HincII (blunts ends) or Pst I (5' recessed ends) in 30 minutes at 37°C. Dephosphorylation is defined as > 95% inhibition of recirculation in a self-ligation reaction and is measured by transformation into *E. coli*.

**Unit Assay Conditions:** Vector DNA is dephosphorylated in restriction endonuclease buffer supplemented with Antarctic Phosphatase Reaction Buffer. Ligation is performed with 50 ng of vector using the NEB Quick Ligation Kit (NEB #M2200).

### Vector Dephosphorylation Protocol:

1. Add 1/10 volume of 10X Antarctic Phosphatase Reaction Buffer to 1–5 µg of DNA cut with any restriction endonuclease in any buffer.
2. Add 1 µl of Antarctic Phosphatase (5 units) and mix.
3. Incubate for 15 minutes at 37°C for 5' extensions or blunt-ends, 60 minutes for 3' extensions.
4. Heat inactivate for 5 minutes at 70°C (or as required to inactivate the restriction enzyme).
5. Proceed with ligation.

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**Usage Notes:** Antarctic Phosphatase is also active in NEBuffers 1, 2, 3 or 4 as well as the NEBuffer for EcoRI **ONLY** when supplemented with 10X Antarctic Phosphatase Reaction Buffer to a final concentration of 1X. Adding 1/10 volume of 10X Antarctic Phosphatase Reaction Buffer will provide the amount of Zn<sup>2+</sup> that the enzyme requires for activity.

### Quality Control Assays

**Exonuclease Activity:** Incubation of a 50 µl reaction containing 50 units of Antarctic Phosphatase with 1 µg of a mixture of single and double-stranded [<sup>3</sup>H] *E. coli* DNA for 4 hours at 37°C released < 0.1% of the total radioactivity.

**Endonuclease Activity:** Incubation of a 50 µl reaction containing 50 units of Antarctic Phosphatase with 1 µg of φX174 RF I DNA for 4 hours at 37°C resulted in < 10% conversion to RFI as determined by agarose gel electrophoresis.

**RNase Activity:** Incubation of a 10 µl reaction containing 5 units of Antarctic Phosphatase with 40 ng of fluorescein labeled RNA transcript for 4 hours at 37°C resulted in no detectable degradation of the RNA as determined by gel electrophoresis using fluorescence detection.

CERTIFICATE OF ANALYSIS

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**Physical Purity:** Purified to > 95% homogeneity as determined by SDS-PAGE analysis using Coomassie Blue detection.

**Transformation Assay:** pUC19 was cleaved with HindIII, HincII or PstI, each purified by Qiaprep™ spin column and resuspended in water at 0.2 mg/ml. 1 µg of each DNA was treated with 5 units of Antarctic Phosphatase. Each reaction was carried out with 1 µg of DNA in a 50 µl reaction volume with 1X Antarctic Phosphatase Reaction Buffer for 30 minutes at 37°C, followed by heat inactivation at 65°C for 5 minutes.

Ligations were performed using the NEB Quick Ligation Kit protocol with 2.5 µl (50 ng) of vector DNA directly from the heat-inactivated phosphatase reaction mix. Inserts were included in 3-fold molar excess. Either HindIII cleaved fragments of λ DNA, HaeIII cleaved fragments of φX174 RF I DNA or PstI cleaved fragments of λ DNA were inserted as indicated.

5 ng of each ligation was transformed into *E. coli* DH5-α. The equivalent of 1.0 ng were plated on LB plates that contained IPTG, X-gal and ampicillin.

Page 2 (M0289)

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	Antarctic Phosphatase Units	Blue Colonies	White Colonies	Background Reduction (%)	Insert (%)
<b>5' Overhang</b>					
HindIII cleaved and ligated	0	1360	0		
Phosphatase treated and ligated	5	16	0		
Phosphatase treated, insert ligated	5	24	216	98.2	15.9
<b>Blunt End</b>					
HincII cleaved and ligated	0	1028	0		
Phosphatase treated and ligated	5	0	0		
Phosphatase treated, insert ligated	5	24	92	97.7	8.9
<b>3' Overhang</b>					
PstI cleaved and ligated	0	872	0		
Phosphatase treated and ligated	5	8	0		
Phosphatase treated, insert ligated	5	20	348	97.7	40.0

**Heat Inactivation:** 70°C for 5 minutes.

**References:**

1. Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, (2nd ed.), (p. 5.72). Cold Spring Harbor: Cold Spring Harbor Laboratory Press.
2. Rina, M. et al. (2000) *Eur. J. Biochem.* 267, 1230–1238.
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