

## New England Biolabs Certificate of Analysis

**Product Name:** *Ph.D.<sup>™</sup>-7 Phage Display Peptide Library Kit*  
**Catalog Number:** *E8100S*  
**Packaging Lot Number:** *10061540*  
**Expiration Date:** *12/2021*  
**Storage Temperature:** *-20°C*  
**Specification Version:** *PS-E8100S v1.0*

Ph.D. <sup>™</sup> -7 Phage Display Peptide Library Kit Component List			
NEB Part Number	Component Description	Lot Number	Individual QC Result
S1259AVIAL	-96 gIII Sequencing Primer (20-mer)	10043734	Pass
S1258AVIAL	-28 gIII Sequencing Primer (22-mer)	10043733	Pass
N7024AVIAL	Biotin	10043732	Pass
N7023AVIAL	Streptavidin, lyophilized	10060978	Pass
E8102AVIAL	Ph.D. <sup>™</sup> -7 Phage Display Peptide Library	10061541	Pass
E4104SVIAL	E.coli K12 ER2738	10034478	Pass

Assay Name/Specification	Lot # 10061540
<b>Sequence Verification (DNA)</b> The Ph.D. <sup>™</sup> -7 Phage Display Peptide Library was sequenced using 5'-CCCATGTACCGTAACACTGAGTTTC-3' as a primer to confirm the correct form of the cloned insert on the displayed peptide, X7-GGG.	<b>Pass</b>
<b>Absolute Phage Titer</b> Infection of a mid-log culture of E. coli ER2738 with Ph.D. <sup>™</sup> -7 Phage Display Peptide Library followed by plating, yields $\geq 1 \times 10^3$ pfu/ml.	<b>Pass</b>
<b>Functional Testing (Panning)</b> A 100-fold representation of the Ph.D. <sup>™</sup> -7 Phage Display Peptide Library containing approximately 1011 pfu is diluted in 200 $\mu$ l TBS and panned against 300 ng $\beta$ -endorphin monoclonal antibody. The bound phage is affinity captured using magnetic beads and eluted with 1 ml of 0.2M Glycine-HCl, pH 2.2. After three rounds of selection, $\geq 75\%$ of sequences contain a motif related to the known epitope for the antibody.	<b>Pass</b>
<b>Phage Contamination (Environmental)</b> A 1:100 dilution of an overnight culture of E. coli ER2738 was made in 20 ml LB, to which $10^5$ pfu of Ph.D. <sup>™</sup> -7 Phage Display Peptide Library was added. The flask was	<b>Pass</b>

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incubated at 37°C on a rotating shaker for 5 hours. A 1 ml volume of culture was removed and centrifuged. Five microliters (5 µl) of phage-containing supernatant was used for three successive rounds of amplification. The final culture supernatant was plated on three LB/IPTG/Xgal plates and then titered. Fewer than 5% clear or white plaques were observed in a minimum of 100 total plaques counted on each plate.	

This product has been tested and shown to be in compliance with all specifications.

*Beth M. Paschal*

Beth Paschal  
Production Scientist  
17 Dec 2019

*Michael Tonello*

Michael Tonello  
Packaging Quality Control Inspector  
18 Dec 2019